

Taku Misonou, Mamiko Seto and Takeshi Nitta: Phylogenetic relationship of three *Bryopsis* species (Derbesiales, Chlorophyta) based on 16S ribosomal RNA sequences

Key Index Words: *Bryopsis* (Chlorophyta)—chloroplast 16S rRNA—molecular evolution—phylogeny—sequence divergence.

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We attempted a molecular phylogenetic approach based on small-subunit rRNA (ribosomal RNA) sequences in three members of genus *Bryopsis*. In this study, we targeted the chloroplast 16S rRNA molecule for the partial sequencing. Their phylogenetic relationship previewed by organelle DNA RFLP (Restriction Fragment Length Polymorphism) analysis (Misonou *et al.* 1989) and some characteristics of *Bryopsis* 16S rRNA were discussed. The algae used were *Bryopsis maxima* Okamura, *B. plumosa* (Hudson) C. Agardh, and *B. sp.* (Misonou *et al.* 1989). The first was collected at Choshi in Chiba, on the Pacific coast, the second at Yokosuka in Kanagawa, and the third at Futtsu in Chiba, from November 1986 to May 1990. The second and third sites are on Tokyo Bay.

Chloroplasts were isolated from algal thalli in an alkaline buffer containing EDTA (Misonou *et al.* 1989, Ishihara *et al.* 1992). Total nucleic acids were extracted by the chloroform/phenol method from this fraction and RNAs were purified with the DNase I treatment. 16S rRNA partial sequences of each species were determined directly with dideoxynucleotide chain termination method using reverse transcriptase according to Lane *et al.* (1985). The three primers used are complementary to the *Escherichia coli* 16S rRNA highly conserved region. Their positions and sequences are I: 357-342 (5'CTGCTGCCTCCCGTAGOH3'), II: 1242-1227 (5'CCATTGTAGCACGTGTOH3') and III: 1510-1495 (5'GGCTACCTTGTTACGAOH3') according to *E. coli* nomenclature

(Sawada *et al.* in press).

The sequences read with each primer were 466 bases and aligned with the homologous sequence of *Chlorella vulgaris* strain 211-11b 16S rRNA (Neefs *et al.* 1990) as an outgroup in Fig. 1. *Bryopsis* 16S rRNAs show intra-generic sequence divergence between each species (Fig. 1). It might be considered that the speciation of these organisms is an ancient incident or the rate of 16S rRNA molecule evolution in this genus is faster than that of higher plant.

Of the total 466 bases, 443 bases were comparable to each other in sequence. The remaining of 23 bases could not be identified by anomalous bands across the lanes on the gel owing to the nucleotide modification. While the sequences with primer II were homologous, the primer I region of *B. plumosa* has 1 deletion (No. 103 in Fig. 1 I), and sequences with primer III were heterogeneous (Fig. 1). In this region, the chain termination reaction continued only up to 100 bases with 19 anomalous bands. It seems that primer III complementary to *E. coli* sequence anneals to *Bryopsis* RNAs in low extent due to the sequence heterogeneity, and/or this region is abundant in modified nucleotides.

The genetic distances of these three regions were calculated according to Kimura's 2 parameter method (Kimura 1980) and presented in Table 1. The primer III region of 3 *Bryopsis* is not only heterogeneous each other but also shows low homology with *Chlorella* sequence. Moreover, no homologous sequence with this region was found in GEN-

I

1
M CCGGCACAGA GUCAGGGUCA CACCAACUAG UAGGAGAGUC UGGUUGAUGA CUAGCUGCGG
S CCGGCACAGA GUCAGGGUCA CACCAACUAG UAGGAGAGUC UGGUUGAUGA CUAGCUGCGG
P CCGGCACAGA GUCAGGGUCA CACCAACUAG UAGGAGAGUC UGGUUGAUGA CUAGCUGCGG
C CCGGCACAGA GUCAGGGUCA CACCGACUAG UAGGAGAGUC UGGUCGAUGA CUAGUAACGG

71
M AACCAUUCGG UAAUGGAGUG GUUGAUCGAU UAGUCUGCGU UCGAGUAAAA AUCCGUCUAG
S AACCAUUCGG UAAUGGAGUG GUUGAUCGAU UAGUCUGCGU UCGAGUAAAA AUCCGUCUAG
P AACCAUUCGG UAAUGGAGUG GUUGAUCGAU UAGUCUGCGU UCGAGUAAAA AUCCGUCUA-
C AACCAUUCGG UAAUGGAGUG GUUGUUCGAU UAGUCGGCGU UCGGGUAGAU AACCGCUAAA

131
M UCUGGAAACU GAAAAGUCG- AAUAAUUCAU AAUCGUUAGC AAAGGUUAAC AAUAAGGAGU
S UCUGGAAACU GAAAAGUCG- AAUAAUUCAU AAUCGUUAGC AAAGGUUAAC AAUAAGGAGU
P UCUGGAAACU GAAAAGUCG- AAUAAUUCAU AAUCGUUAGC AAAGGUUAAC AAUAAGGAGU
C AGUAGAAAGU GAAGAGUCGU UAUGCUCUAA AAUCGUUAGC AAAGGUUACC AAUAGGGAGA
..***
191
M GGAUUUACAU CUAAGAAUGC GCAAUGAGUGG
S GGAUUUACAU CUAAGAAUGC GCAAUGAGUGG
P GGAUUUACAU CUAAGAAUGC GCAAUGAGUGG
C GGUUUUCCA CCAAGAAUGC ACAAUGAGUGG
**.*.*.*.*

II

1
M CCCGAAUUC CCGUACGACU GAACUGCAGU AGGAGUGGAA GGAGGCCAAA UAGUGGCGGU
S CCCGAAUUC CCGUACGACU GAACUGCAGU AGGAGUGGAA GGAGGCCAAA UAGUGGCGGU
P CCCGAAUUC CCGUACGACU GAACUGCAGU AGGAGUGGAA GGAGGCCAAA UAGUGGCGGU
C CCUGCAUUC CCGUACGACU GAACUGCAGU AGGAGUGGAA GGAGGCCGAA CAGUGGCCGU
**.*.*.*.*
71
M CAGAGAGAUC UAU---UAAU -----UGA-U UUCUGUCC C AACGCGAGCA ACXCC-UGAX
S CAGAGAGAUC UAU---UAAU -----UGA-U UUCUGUCC C AACGCGAGCA ACXCC-UGAX
P CAGAGAGAUC UAU---UAAU -----UGA-U UUCUGUCC C AACGCGAGCA ACXCC-UGAX
C CAGAAACU AAAGGUAAU GACCGUUAAG UUUUGUCC C AACGCGAGCA ACGCCUGAA
***.*.*.*.*
131
M UUGGGUUGUA GAGUGCUGUX XUCGACUGCU GUCGGUACGU GGUGGA
S UUGGGUUGUA GAGUGCUGUX XUCGACUGCU GUCGGUACGU GGUGGA
P UUGGGUUGUA GAGUGCUGUX XUCGACUGCU GUCGGUACGU GGUGGA
C UUGGGUUGUA GAGUUCUGUG CUCGACUGCU GUCGCUACGU GGUGGA

III

1
M GGCUXAGCUG GUUCCGGUGG CCUCXACCXU UXGGCCGGCC CUCCGUCC C CGGUUXCGXX
S GGCUXAGCUG GUUCCGGGGG CCUCXACCXU UXGGCCGGCC CUCCGUCC C CGGUUXCGXX
P GGCUXAGCUG GUUCCGGUGG CCUCXACCXU UXGGCCGGCC CUCCGUCC C CGGCUXCGXX
C GGAUCAG-UG AUCGAGACGG AAUCCGAG- -GGGAGGAU UUCAACCC C --AUUGCUGA
***.*.*.*.*
71
M XACUCXCCCC --UCAGXAUX GCXACGGXGU GXGX---ACA XXXXC
S XACUCXCCCC --UCAGXAUX GCXACGGXGU GXGX---ACA XXXXC
P XACUCXCCCC --UCAGXAUX GCXACGGXGU GXGX---ACA XXXXC
C AACCCGUAUC GGUCGAGGGU ACCACACUGC CCGCCACAGA UGUUC
**.*.*.*.*

Table 1. Genetic distances ($\times 10^{-3}$ Knuc) of three regions between each of three *Bryopsis* and *Chlorella* 16S rRNA.

region	M-S	M-P	S-P	M-C	S-C	P-C
I	0.0	0.0	0.0	171.4	171.4	166.1
II	0.0	0.0	0.0	113.5	113.5	113.5
III	12.5	25.1	12.3	681.5	673.0	756.5
total	2.3	4.5	6.8	213.9	213.7	217.4

M: *Bryopsis maxima*; S: *B. sp.*; P: *B. plumosa*; C: *Chlorella vulgaris*.

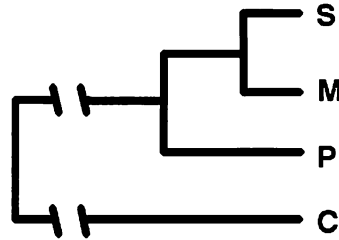
BANK database. These results show that *Bryopsis* 16S RNA sequences around the position 1400–1500 are not conservative even in the primer III region considered homologous from bacteria to higher plant chloroplast. This may suggest that these algae occupy a unique position in plant kingdom.

From these genetic distances, a phylogenetic tree was inferred in Fig. 2 by UPGMA (Sokal and Michener 1958), and NJ method (Saitou and Nei 1987) with bootstrap confidence limits using the Clustal V computer program (Higgins 1991). In both trees, *B. sp.* is distinguished from other 2 species. It is shown that this alga, having regarded as an intraspecific variation of *B. plumosa*, is an independent species and has rather far relationship with *B. plumosa*. In the UPGMA tree, *B. sp.* is clustered with *B. maxima* as previous RFLP analysis (Misonou *et al.* 1989). On the other hand, although the bootstrap confidence interval is not sufficiently high, *B. sp.* is separated from *B. maxima*—*B. plumosa* cluster in the NJ tree. In the case of this study, UPGMA tree seems to be more reliable for the close relationship of these algae. More sequence informations may be required for further phylogenetic analyses of *Bryopsis*.

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UPGMA



NJ

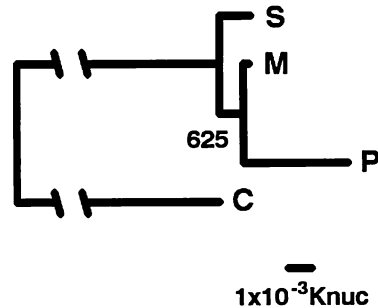


Fig. 2. Phylogenetic trees drawn with genetic distances of three *Bryopsis* and *Chlorella* 16S rRNA partial sequences using 2 tree construction methods. M, *Bryopsis maxima*; S, *B. sp.*; P, *B. plumosa*; C, *Chlorella vulgaris* 211–11b. The number under the branch in NJ tree is the confidence interval for nodes based upon 1,000 bootstrap samples.

ful suggestion pertaining to *Bryopsis* classification. We also thank Prof. K. Wakabayashi, Yamanashi Medical College for his kind help to this study.

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References

- Higgins, D. G. 1991. Clustal V Documentation. EMBL.
- Ishihara, J., Pak, J. Y., Fukuhara, T. and Nitta, T. 1992. Association of particles that contain double-stranded RNAs with algal chloroplasts and mitochondria. *Planta* **187**: 475–482.
- Kimura, M. 1980. A simple method for estimating evolutionary rates of base substitutions through com-

Fig. 1. Partial sequences of three *Bryopsis* 16S rRNA. The sequences determined with three primers are aligned. M, *Bryopsis maxima*; S, *B. sp.*; P, *B. plumosa*; C, *Chlorella vulgaris* 211–11b. Hyphens and Xs show deletions and unclotides that could not be identified, respectively. Asterisks are the homologous nucleotides with three *Bryopsis* and *Chlorella*. Dots are the nucleotides homologous within three *Bryopsis* while not with *Chlorella*.

- parative studies of nucleotide sequences. *J. Mol. Evol.* **16**: 111-120.
- Lane, J., Pace, B., Olsen, G. J., Stahl, D. A., Sogin, M. L. and Pace, N. R. 1985. Rapid determination of 16S ribosomal RNA sequences for phylogenetic analyses. *Proc. Natl. Acad. Sci. USA* **82**: 6955-6959.
- Misonou, T., Ishihara, J., Pak, J. Y. and Nitta, T. 1989. Restriction endonuclease analysis of chloroplast and mitochondrial DNAs from *Bryopsis* (Derbesiales, Chlorophyta). *Phycologia* **28**: 422-428.
- Neefs, J.-M., Van de Peer, Y., Hendriks, L. and De Wachter, R. 1990. Compilation of small ribosomal subunit RNA sequences. *Nuc. Acid Res.* **18**: 2237-2317.
- Saitou, N. and Nei, M. 1987. The neighbor-joining method: A new method for reconstructing phylogenetic trees. *Mol. Biol. Evol.* **4**: 406-425.
- Sawada, H., Ieki, H., Oyaizu, H. and Matsumoto, S. 1993. Proposal for rejection of *Agrobacterium tumefaciens* and revised descriptions for the genus *Agrobacterium* and *Agrobacterium radiobacter* and *Agrobacterium rhizogenes*. *Int. J. Syst. Bacteriol.* (in press)
- Sokal, R. R. and Michener, C. D. 1958. A statistical method for evaluating systematic relationships. *Univ. Kansas Sci. Bull.* **28**: 1409-1438.

御園生 拓*・瀬戸麻美子*・新田 毅** : 16S リボソーム RNA 塩基配列による
3 種のハネモ属藻 (ツクノイト目, 緑藻植物門) の系統関係

大型緑藻ハネモ属の系統を明らかにするために、葉緑体 16S rRNA (リボソーム RNA) の塩基配列を比較し解析することを試みた。逆転写酵素を利用したサンガー反応のプライマーには 16S rRNA に特異的な保存配列を 3 種類用い、計 466 塩基を読んだ。ミナトハネモ (*Bryopsis* sp.) は独立した種であり、ハネモ (*B. plumosa*) やオオハネモ (*B. maxima*) とは遺伝的に離れているという結果を得た。また、これらのハネモ 16S rRNA の塩基 No. 1400~1500 に相当する部分は既知の生物の配列との相同性が低く、これらが他の植物と系統的に離れている可能性が示唆された。(*400 甲府市武田 4 山梨大学教育学部生物学教室, **183 府中市幸町 3 東京農工大学一般教育部生物)

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